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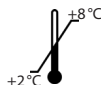
IDK[®] Calprotectin ELISA

***For the in vitro determination of calprotectin (MRP 8/14,
S100A8/A9) in serum and plasma***

Valid from 2022-03-22



K 6935



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1. INTENDED USE

This Immundiagnostik AG assay is an enzyme immunoassay intended for the quantitative determination of calprotectin (MRP 8/14, S100A8/A9) in serum and plasma. For *in vitro* diagnostic use only.

2. INTRODUCTION

Alternative names of calprotectin:

- MRP8/14, L1, (p8,14), p34, S100 A8/A9

Alternative names of the two proteins forming the heterocomplex calprotectin:

- S100A8, Calgranulin A, MRP8 (Migration inhibition factor-related protein-8), CP-10 (in mouse)
- S100A9, Calgranulin B, MRP14 (Migration inhibition factor-related protein-14)

Calprotectin is a calcium-binding protein secreted predominantly by neutrophils and monocytes. The heterocomplex consists of the two proteins, S100A8 (calgranulin A) and S100A9 (calgranulin B), also designated as MRP8 and MRP14, respectively. Expression of S100A8 and S100A9 in epithelial tissues was first described in context with squamous epithelia and with murine and human wound repair. More recently, an association of S100 protein expression with adenocarcinomas in humans has emerged. The genes S100A8 and S100A9 are located in a gene cluster on chromosome 1q21, a region in which several rearrangements that occur during tumor development have been observed.

Elevated MRP8/14 levels have been found in many sites of inflammation and in the extracellular fluid of patients with many types of inflammatory conditions. The concentration of MRP8/14 in blood is increased in patients with rheumatoid arthritis, cystic fibrosis, multiple sclerosis, and HIV infections, while elevated MRP8/14 levels have been detected in stool of patients with Crohn's disease and colorectal cancer [1-5]. Extracellular MRP8/14 has antimicrobial, antigrowth and apoptotic effects. It suppresses the growth of some fungi and bacteria [1,2]. It also suppresses the proliferation of several different types of cells including: macrophages, lymphocytes, hematopoietic progenitors, and tumor cell lines. MRP8/14 can also induce apoptosis of some tumor cell lines [1,2].

Hermani et al. (2005) [6] reported that enhanced expression of S100A8 and S100A9 is an early event in prostate tumor genesis and may contribute to development and progression or extension of prostate carcinomas. Furthermore, they tested the value of S100A9 as a serum marker for prostate cancer comparing the serum concentrations of S100A9 in cancer patients with healthy controls or patients with benign prostatic hyperplasia (BPH). Significantly increased S100A9 serum levels in prostate

cancer were found in prostate cancer patients compared to patients with BPH, the latter exhibiting values similar to that obtained for healthy individuals.

Pathological significance and clinical application

The diagnostic value and advantage of MRP8/14 over other disease markers is that they are preformed and released immediately upon activation of the respective cell population. Other markers may be generated in downstream events or need to be synthesised de novo in the liver. Various conditions have shown significant correlation of MRP8/14 (or MRP8, MRP14) levels with disease activity:

- Concentrations of MRP8/14 in serum, and particularly in synovial fluid, correlate strongly with disease activity in rheumatoid arthritis.
- Plasma MRP8/14 levels are very early, specific and sensitive prediction markers for acute rejection in kidney allograft transplantation.
- Serum MRP8/14 concentration is a prognostic marker of recurrent infection and survival in alcoholic liver cirrhosis.
- MRP8/14 is useful for evaluating the extent of periodontal inflammation.
- In cerebral malaria, MRP 8/14 expression correlates with microglial activation in brain.
- MRP8/14 is present in urinary stones and in dental calculus.
- S100A9 in serum may serve as a useful marker for discrimination between prostate cancer and benign prostatic hyperplasia (BPH).

3. MATERIAL SUPPLIED

Cat. No.	Label	Kit components	Quantity
K 6935	PLATE	Microtiter plate, pre-coated	12 x 8 wells
K 0001.C.100	WASHBUF	Wash buffer concentrate, 10x	2 x 100 ml
K 6935	SAMPLEBUF	Sample dilution buffer, ready-to-use	1 x 100 ml
K 6935	STD	Calprotectin standards, ready-to-use (0; 3,9; 15,6; 62,5; 250 ng/ml)	1 x 5 vials
K 6935	CTRL 1	Control, ready-to-use (see specification for range)	1 x 1 vial
K 6935	CTRL 2	Control, ready-to-use (see specification for range)	1 x 1 vial
K 6935	CONJ	Conjugate, ready-to-use	1 x 15 ml
K 0002.15	SUB	Substrate (tetramethylbenzidine), ready-to-use	1 x 15 ml

Cat. No.	Label	Kit components	Quantity
K 0003.15	STOP	Stop solution, ready-to-use	1 x 15 ml

For reorders of single components, use the catalogue number followed by the label as product number.

4. MATERIAL REQUIRED BUT NOT SUPPLIED

- Ultrapure water*
- Calibrated precision pipettors and 10–1 000 µl single-use tips
- Foil to cover the microtiter plate
- Multi-channel pipets or repeater pipets
- Vortex
- Standard single-use laboratory glass or plastic vials, cups, etc.
- Microtiter plate reader (required filters see chapter 7)

* Immundiagnostik AG recommends the use of ultrapure water (water type 1; ISO 3696), which is free of undissolved and colloidal ions and organic molecules (free of particles > 0.2 µm) with an electrical conductivity of 0.055 µS/cm at 25 °C (≥ 18.2 MΩ cm).

5. PREPARATION AND STORAGE OF REAGENTS

- To run the assay more than once, ensure that reagents are stored at the conditions stated on the label. **Prepare only the appropriate amount necessary for each run.** The kit can be used up to 4 times within the expiry date stated on the label.
- **Preparation of the wash buffer:** The **wash buffer concentrate (WASHBUF)** has to be diluted with ultrapure water **1:10** before use (100 ml WASHBUF + 900 ml ultrapure water), mix well. Crystals could occur due to high salt concentration in the concentrate. Before dilution, the crystals have to be redissolved at room temperature or in a water bath at 37 °C. The **WASHBUF** is stable at **2–8 °C** until the expiry date stated on the label. **Wash buffer** (1:10 diluted WASHBUF) can be stored in a closed flask at **2–8 °C for 1 month**.
- All other test reagents are ready-to-use. Test reagents are stable until the expiry date (see label) when stored at **2–8 °C**.

6. STORAGE AND PREPARATION OF SAMPLES

Sample stability and storage

Calprotectin is stable in serum for 7 days at 2–8°C as well as for 3 days at room temperature. At -20°C, the samples can be stored for up to 6 months. More than 3 freeze-thaw cycles are to be avoided.

Calprotectin is not stable in plasma.

Preanalytic handling

Significant differences in the calprotectin levels can be observed due to different sample preparation procedures, e. g. up to 10-fold higher serum levels compared to the plasma calprotectin concentrations. The reasons are as follows:

Granulocytes are activated during serum clotting and release granulocyte-activating markers. The time between serum collecting and analysis as well as repeated freeze-thaw cycles don't cause a calprotectin concentration shift.

On the contrary, in the case of plasma samples, varying the time between sampling and analysis or the number of freeze-thaw cycles will cause variation in the observed calprotectin levels. Therefore, the preanalytical conditions of plasma samples should be held constant. This is a general requirement independent of the used test-system.

Immundiagnostik AG recommends the use of serum samples for calprotectin determinations.

Lipemic or hemolytic samples may give erroneous results and should not be used for analysis.

Sample preparation

Serum samples

Serum samples must be diluted **1:100 with sample dilution buffer** (SAMPLEBUF) before performing the assay, e. g.

50 µl sample + 450 µl SAMPLEBUF = dilution I (1:10)

50 µl dilution I + 450 µl SAMPLEBUF = dilution II (1:10)

Final dilution 1:100

For analysis, pipet **100 µl of dilution II** per well.

Plasma samples

EDTA plasma samples must be diluted **1:30 with sample dilution buffer** (SAMPLE-BUF) before performing the assay, e. g.

20 µl sample + **580 µl** (SAMPLEBUF).

For analysis, pipet **100 µl** of the **dilution** per well.

7. ASSAY PROCEDURE

Principle of the test

This ELISA is designed for the quantitative determination of calprotectin (MRP (8/14, S100A8/A9). The assay utilises the two-site sandwich technique with two selected monoclonal antibodies that bind to human calprotectin.

Standards, controls and diluted patient samples which are assayed for human calprotectin are added to wells of microplate coated with a high affine monoclonal anti-human calprotectin antibody. During the first incubation step, calprotectin in the samples is bound by the immobilised antibody. Then a peroxidase labelled conjugate is added to each well and the following complex is formed: capture antibody - human calprotectin – peroxidase conjugate. Tetramethylbenzidine (TMB) is used as a substrate for peroxidase. Finally, an acidic stop solution is added to terminate the reaction. The colour changes from blue to yellow. The intensity of the yellow colour is directly proportional to the calprotectin concentration of sample.

A dose response curve of the absorbance unit (optical density, OD at 450 nm) vs. concentration is generated, using the values obtained from standard. Calprotectin, present in the patient samples, is determined directly from this curve.

Test procedure

Bring all **reagents and samples to room temperature** (15–30 °C) and mix well.

Mark the positions of standards/controls/samples on a protocol sheet.

Take as many **microtiter strips** as needed from kit. Store unused strips together with the desiccant bag in the closed aluminium packaging at 2–8 °C. Strips are stable until expiry date stated on the label.

For automated ELISA processors the given protocol may need to be adjusted according to the specific features of the respective automated platform. For further details please contact your supplier or Immundiagnostik AG.

We recommend to carry out the tests in duplicate.

1.	Add each 100 µl standards/controls/diluted samples into the respective wells.
2.	Cover the strips and incubate for 30 min at room temperature (15–30 °C).
3.	Discard the content of each well and wash 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
4.	Add 100 µl conjugate (CONJ) into each well.
5.	Cover the strips and incubate for 30 min at room temperature (15–30 °C).
6.	Discard the content of each well and wash 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
7.	Add 100 µl substrate (SUB) into each well.
8.	Incubate for 10–20 minutes* at room temperature (15–30 °C) in the dark .
9.	Add 100 µl stop solution (STOP) into each well and mix well.
10.	Determine absorption immediately with an ELISA reader at 450 nm against 620 nm (or 690 nm) as a reference. If no reference wavelength is available, read only at 450 nm. If the extinction of the highest standard exceeds the range of the photometer, absorption must be measured immediately at 405 nm against 620 nm as a reference.

* The intensity of the colour change is temperature sensitive. We recommend to observe the colour change and to stop the reaction upon good differentiation.

8. RESULTS

The following algorithms can be used alternatively to calculate the results. We recommend using the „4 parameter algorithm“.

1. 4 parameter algorithm

It is recommended to use a linear ordinate for the optical density and a logarithmic abscissa for the concentration. When using a logarithmic abscissa, the zero calibrator must be specified with a value less than 1 (e.g. 0.001).

2. Point-to-point calculation

We recommend a linear ordinate for the optical density and a linear abscissa for the concentration.

3. Spline algorithm

We recommend a linear ordinate for the optical density and a linear abscissa for the concentration.

The plausibility of the pairs of values should be examined before the automatic evaluation of the results. If this option is not available with the used program, a control of the paired values should be done manually.

Serum

The obtained results have to be multiplied by the **dilution factor of 100** to get the actual concentrations.

In case **another dilution factor** has been used, multiply the obtained result by the dilution factor used.

EDTA plasma

The obtained results have to be multiplied by the **dilution factor of 30** to get the actual concentrations.

In case **another dilution factor** has been used, multiply the obtained result by the dilution factor used.

9. LIMITATIONS

Samples with concentrations above the measurement range (see definition below) can be further diluted and re-assayed. Please consider this higher dilution when calculating the results.

Samples with concentrations lower than the measurement range (see definition below) cannot be clearly quantified.

The upper limit of the measurement range can be calculated as:

highest concentration of the standard curve × sample dilution factor to be used

The lower limit of the measurement range can be calculated as:

LoQ × sample dilution factor to be used

LoQ see chapter "Performance Characteristics".

10. QUALITY CONTROL

Immundiagnostik AG recommends the use of external controls for internal quality control, if possible.

Control samples should be analysed with each run. Results, generated from the analysis of control samples, should be evaluated for acceptability using appropriate statistical methods. The results for the patient samples may not be valid if within the same assay one or more values of the quality control sample are outside the acceptable limits.

Reference range

Calprotectin in serum of healthy persons: < 3 µg/ml (< 3 000 ng/ml)

We recommend each laboratory to establish its own reference concentration range.

11. PERFORMANCE CHARACTERISTICS

Accuracy – Precision

Repeatability (Intra-Assay); n = 80

The repeatability was assessed with 2 serum samples under **constant** parameters (same operator, instrument, day and kit lot).

Sample	Mean value [ng/ml]	CV [%]
1	447.7	3.0
2	788.8	3.3

Reproducibility (Inter-Assay); n = 27

The reproducibility was assessed with 3 serum samples under **varying** parameters (different operators, instruments, days and kit lots).

Sample	Mean value [ng/ml]	CV [%]
1	484.7	8.6
2	649.8	9.0
3	827.2	7.6

Accuracy – Trueness

The trueness states the closeness of the agreement between the result of a measurement and the true value of the measurand. Therefore, calprotectin spikes with known concentrations were added to 2 different serum samples. The results below were obtained without consideration of the sample dilution factor.

Sample [ng/ml]	Spike [ng/ml]	Expected [ng/ml]	Obtained [ng/ml]	Recovery [%]
4.4	8.4	12.7	11.4	90.0
	13.2	17.4	16.4	94.1
	16.8	21.0	19.3	91.8
	24.1	28.3	27.2	96.3
7.5	8.4	15.7	15.4	98.1
	13.2	20.5	20.3	99.0
	16.8	24.0	24.0	99.7
	24.1	31.2	30.3	97.0

Linearity

The linearity states the ability of a method to provide results proportional to the concentration of analyte in the test sample within a given range. This was assessed according to CLSI guideline EP6-A with a serial dilution of 2 different serum samples.

For calprotectin in serum and plasma, the method has been demonstrated to be linear from 6.0 to 89.1 ng/ml based on the standard curve without considering possibly used sample dilution factors, showing a non-linear behaviour of less than $\pm 20\%$ in this interval.

Sample	Dilution	Expected [ng/ml]	Obtained [ng/ml]	Recovery [%]
A	1:100	89.1	89.1	100.0
	1:200	44.5	45.2	101.4
	1:400	22.3	18.9	85.0
	1:800	11.1	9.2	82.7

Sample	Dilution	Expected [ng/ml]	Obtained [ng/ml]	Recovery [%]
B	1:100	61.6	61.6	100.0
	1:200	30.8	30.0	97.4
	1:400	15.4	13.7	88.8
	1:800	7.7	6.0	78.4

Analytical sensitivity

The following values have been estimated based on the concentrations of the standard without considering possibly used sample dilution factors.

Limit of blank, LoB	0.522 ng/ml
Limit of detection, LoD	0.789 ng/ml
Limit of quantitation, LoQ	0.897 ng/ml

The evaluation was performed according to the CLSI guideline EP17-A2. The specified accuracy goal for the LoQ was 20% CV.

Analytical specificity

The specificity of the antibody was tested by measuring the cross-reactivity against a range of compounds with structural similarity to calprotectin. There was no cross-reactivity observed.

Substance tested	Concentration added [ng/ml]	Concentration obtained [ng/ml]	Conclusion
Lysozyme	10 000	< 0.522	< LoB
PMN elastase	1 000	2.003	0.2 %
MPO	10 000	< 0.522	< LoB
Lactoferrin	100 000	< 0.522	< LoB

Traceability

The standards of the IDK® Calprotectin ELISA are based on recombinant human calprotectin which has been calibrated against purified MRP8/14 derived from human granulocytes.

12. PRECAUTIONS

- All reagents in the kit package are for *in vitro* diagnostic use only.
- Human materials used in kit components were tested and found to be negative for HIV, Hepatitis B and Hepatitis C. However, for safety reasons, all kit components should be treated as potentially infectious.
- Kit reagents contain sodium azide or ProClin as bactericides. Sodium azide or ProClin are hazardous to health and the environment. Substrates for enzymatic colour reactions may also cause skin and/or respiratory irritation. Any contact with the substances must be avoided. Further safety information can be found in the safety data sheet, which is available from Immundiagnostik AG on request.
- The 10x Wash buffer concentrate (WASHBUF) contains surfactants which may cause severe eye irritation in case of eye contact.

Warning: Causes serious eye irritation. **IF IN EYES:** Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. If eye irritation persists: get medical Advice/attention.

- The stop solution consists of diluted sulphuric acid, a strong acid. Although diluted, it still must be handled with care. It can cause burns and should be handled with gloves, eye protection, and appropriate protective clothing. Any spill should be wiped out immediately with copious quantities of water. Do not breath vapour and avoid inhalation.

13. TECHNICAL HINTS

- Do not interchange different lot numbers of any kit component within the same assay. Furthermore we recommend not to assemble wells of different microtiter plates for analysis, even if they are of the same batch.
- Control samples should be analysed with each run.
- Reagents should not be used beyond the expiration date stated on kit label.
- Substrate solution should remain colourless until use.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Avoid foaming when mixing reagents.
- Do not mix plugs and caps from different reagents.
- The assay should always be performed according to the enclosed manual.

14. GENERAL NOTES ON THE TEST AND TEST PROCEDURE

- This assay was produced and distributed according to the IVD guidelines of 98/79/EC.
- Quality control guidelines should be followed.
- IDK® is a trademark of Immundiagnostik AG.
- Incubation time, incubation temperature and pipetting volumes of the components are defined by the producer. Any variation of the test procedure, which is not coordinated with the producer, may influence the results of the test. Immundiagnostik AG can therefore not be held responsible for any damage resulting from wrong use.
- Warranty claims and complaints in respect of deficiencies must be logged within 14 days after receipt of the product. The product should be sent to Immundiagnostik AG along with a written complaint.

15. REFERENCES

General literature

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Used symbols:

	Temperature limitation		Catalogue number
	In Vitro Diagnostic Medical Device		To be used with
	Manufacturer		Contains sufficient for <n> tests
	Lot number		Use by
	Attention		Consult instructions for use
	Consult specification data sheet		Irritant